

This article was downloaded by: [Politechnika Warszawska]

On: 08 November 2013, At: 05:26

Publisher: Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



High Pressure Research: An International Journal

Publication details, including instructions for authors and subscription information:

<http://www.tandfonline.com/loi/ghpr20>

The sound velocity measurement in diacylglycerol oil under high pressure

A. J. Rostocki^a, A. Malanowski^a, R. Tarakowski^a, K. Szlachta^a, P. Kiełczyński^b, M. Szalewski^b, A. Balcerzak^b & S. Ptasznik^c

^a Faculty of Physics, Warsaw University of Technology, Koszykowa 75, 00-662, Warsaw, Poland

^b Institute of Fundamental Technological Research, Polish Academy of Sciences, Pawińskiego 5B, 02-106, Warsaw, Poland

^c Department of Meat and Fat Technology, Institute of Agricultural and Food Biotechnology, Jubilerska 4, 04-190, Warsaw, Poland

Published online: 25 Feb 2013.

To cite this article: A. J. Rostocki, A. Malanowski, R. Tarakowski, K. Szlachta, P. Kiełczyński, M. Szalewski, A. Balcerzak & S. Ptasznik (2013) The sound velocity measurement in diacylglycerol oil under high pressure, High Pressure Research: An International Journal, 33:1, 172-177, DOI: [10.1080/08957959.2013.769975](https://doi.org/10.1080/08957959.2013.769975)

To link to this article: <http://dx.doi.org/10.1080/08957959.2013.769975>

PLEASE SCROLL DOWN FOR ARTICLE

Taylor & Francis makes every effort to ensure the accuracy of all the information (the "Content") contained in the publications on our platform. However, Taylor & Francis, our agents, and our licensors make no representations or warranties whatsoever as to the accuracy, completeness, or suitability for any purpose of the Content. Any opinions and views expressed in this publication are the opinions and views of the authors, and are not the views of or endorsed by Taylor & Francis. The accuracy of the Content should not be relied upon and should be independently verified with primary sources of information. Taylor and Francis shall not be liable for any losses, actions, claims, proceedings, demands, costs, expenses, damages, and other liabilities whatsoever or howsoever caused arising directly or indirectly in connection with, in relation to or arising out of the use of the Content.

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden. Terms &

Conditions of access and use can be found at <http://www.tandfonline.com/page/terms-and-conditions>

The sound velocity measurement in diacylglycerol oil under high pressure[†]

A.J. Rostocki^{a*}, A. Malanowski^a, R. Tarakowski^a, K. Szlachta^a, P. Kietczyński^b, M. Szalewski^b,
A. Balcerzak^b and S. Ptasznik^c

^aFaculty of Physics, Warsaw University of Technology, Koszykowa 75, 00-662 Warsaw, Poland; ^bInstitute of Fundamental Technological Research, Polish Academy of Sciences, Pawińskiego 5B, 02-106 Warsaw, Poland; ^cDepartment of Meat and Fat Technology, Institute of Agricultural and Food Biotechnology, Jubilerska 4, 04-190 Warsaw, Poland

(Received 28 September 2012; final version received 21 January 2013)

In this article, the influence of high pressure on sound velocity at 293 K has been presented. The investigated diacylglycerol oil (DAG – [D82T18]AG) was composed of 82% DAGs and 18% triacylglycerols. The variation of sound velocity with hydrostatic pressure for DAG was evaluated up to 400 MPa. The phase transformation in DAG has been observed as a discontinuity of the dependence of sound velocity on pressure. The sound velocity during the phase transition has shown distinct increment. Also the volume changes have been measured. It has shown the rapid drop of the volume at the phase transformation pressure due to the possible crystallization of DAG oil.

Keywords: high pressure; velocity; DAG

1. Introduction

Research on the high pressure properties of a wide class of lipid compounds have been stimulated by the development of industrial methods of high pressure food conservation. A special attention was paid to the pressure-induced changes of edible oils since the phase transitions were observed during these processes [1–8]. Natural oils such as castor oil [1], rapeseed oil [2], soy oil [3] and others [5–8] and their main components *i.e.* triacylglycerols (TAGs) [6,9,10] are intensively studied. Later, the research was extended to fatty acids [11–15] and lately on diacylglycerols (DAGs) [16,17]. In the meantime, the range of experimental methods was extended from basic optical measurements [9,10,12,16] to dielectric properties [13] and on ultrasonic methods of measuring viscosity [17,18] and sound velocity [19,20]. In this article, the results of sound velocity measurement during DAG oil compression has been presented.

DAGs are esters of the trihydric alcohol glycerol in which two of the hydroxyl groups are esterified with fatty acids. They consist of two fatty acid chains bond to a glycerol molecule.

*Corresponding author. Email: arostock@if.pw.edu.pl

[†]This article was presented at the Lth European High Pressure Research Group (EHPRG 50) Meeting at Thessaloniki (Greece), 16–21 September 2012.

Table 1. Main components of the investigated DAG oil sample.

Fatty acid carbon atoms: C=C bonds	Amounts (%)
C14:0	0.1
C16:0	4.5
C16:1(Σ)	0.5
C17:0	0.1
C17:1	0.1
C18:0	2.5
C18:1(Σc)	59.1
C18:2(cc)	19.6
C18:3(ccc)	8.9
C20:0	1.9
C20:1(Σ)	1.0
C22:0	0.1
C22:1(σ)	1.2
C24:0	0.3
C24:1	0.1

Since two fatty acid chains can bond to two of the three possible positions, there are three isomers of DAGs named sn-1,2, sn-2,3 and sn-1,3. The isomer sn-1,3 has one additional plane of symmetry, therefore, it has different thermodynamic properties from the other two (for instance higher melting temperature, higher viscosity coefficient). Main components of the investigated sample DAG oil are given in Table 1. Similarly to other lipids, DAG can crystallize into several polymorphic forms [21,22]. In case of natural DAGs, two acyl chains may consist of two different fatty acids of different chain length (the so-called mixed chain DAGs). The most stable crystalline phase is the so-called beta phase, a monoclinic crystalline form with an orthorhombic perpendicular sub-cell chain packing, in which both acyl chains are parallel to each other. The molecules are organized into a bi-layer structure, with the glycerol backbone roughly parallel to the plane of it, and the acyl chains tilted at approximately 60 degrees with respect to that plane. The acyl chain unsaturation, and particularly a single *cis* unsaturation, impairs chain packing. The diversity of the mixed chain DAG results in an increased number of metastable crystalline phases.

Monoacylglycerols and DAGs are common food additives used to blend together certain ingredients, such as oil and water, which would not otherwise blend well. Partially hydrogenated soya bean and canola oil are the main sources of glycerols. They may also be synthetically produced. Edible oils consisting of about 80% of 1,3-DAGs are sold in Japan as nutritional supplements. It is claimed that they are metabolized in a different way than the DAGs are, and with a beneficial nutritional effect [23–26]. The 1(3)-monoacylglycerols when digested are rather poorly absorbed, apparently limiting the accumulation of fats in body tissues.

When fatty materials are used as food products, knowledge of their physical properties is of great importance, especially with regard to interfacial tension, solubility, viscosity, melting behaviour and other physicochemical characteristics of the material. DAG oil produced for direct consumption was investigated in our first experiments.

2. Experimental setup

High pressure measurements were performed using a set-up that was designed and made at Warsaw University of Technology. Pressure was applied by a manually driven hydraulic press [8,27]. Pressure and temperature sensors were located inside the 22 cm³ pressure chamber within the investigated DAG oil sample. The pressure was measured by a manganin sensor, calibrated using a dead weight piston gauge [28]. The manganin gauge's electrical resistance, that is, proportional

to the pressure, was measured by a linear unbalanced bridge [29]. Volume changes were calculated from the piston displacement measured by a digital caliper. Corrections related to the expansion of the chamber were evaluated from the Lamé equations. Temperature was kept constant by a thermostatic water bath at 293 K. The sample density under normal conditions was determined by weighting known volume of the sample. Since the mass of the sample was not changed during the experiment, the variation of the volume allowed determining the density.

Sound velocity was determined based on time of flight of ultrasonic waves between transducers. Two 5 MHz LiNbO₃ (Y36 cut) plates (Boston Piezo-Optics Inc., USA) were used as the transmitter and the receiver. The ultrasonic transducers' mounting has been designed to provide a low level of parasitic ultrasonic signals. The TB-1000 pulser–receiver computer card (Matec, USA) was a signal source for the transmitter. Having passed through the investigated sample, the signal was detected by the receiver and was then processed by the PDA-1000 digitizer card (Signatec, USA). To increase the signal-to-noise ratio, each single measurement was repeated 1024 times and finally averaged. The cross-correlation method [30] was used to evaluate the time of flight. The experimental set-up and method are presented in detail in [19,20]. The pressure was increased in 10 MPa steps and with a time interval which allowed the sample to reach the thermodynamic equilibrium. At a pressure of 210 MPa (at 293 K), a longer relaxation time was used, because at this point the phase transition was found. Subsequently, the pressure was again raised in steps up to 500 MPa and finally it was slowly reduced.

3. Investigated sample

Precise analysis of the DAG sample composition was done at the Laboratory of Institute of Agricultural and Food Biotechnology (Department of Meat and Fat Technology) using the Gas Chromatography method. For this purpose, a HP6890 device was applied following the AOCS Cd 11b-91 official method. The analysis was performed according to ISO 5508 and ISO 5509 norms, whereas the data were analysed with ChemStation a 03.34 software. The main components of the investigated DAG are given in Table 1.

Apart from the main components given in Table 1, the slight amounts of other fatty acids at the level of about 0.1%, were presented in the sample, *i.e.* C14:0, C17:0, C17:1, C22:0 and C24:1. But their influence on the observed effects was considered to be negligible.

4. Results and discussion

During the experiment, the volume variations and sound velocity were measured simultaneously over a pressure range. It was noticed that at a pressure about 210 MPa, the dependence of sound velocity versus pressure has a discontinuity that is characteristic for the first-order phase transition. Above the transition pressure, the observed dependence of volume on pressure is very similar to that observed in triolein [4]. The changes in sound velocity caused by the increase of pressure are shown in Figure 1. At a phase-transition pressure, the discontinuous increase of the speed of sound was noticed, despite the drop of pressure. Similar phenomena were observed in all previously studied triglycerides [19,20]. At a pressure of 400 MPa, the speed of sound is almost twice greater compared to its value at atmospheric pressure. Such value, *i.e.* about 3000 m/s, is typical for solids, which supports the hypothesis that the crystallization process of triglycerides observed in model triglycerides triolein and trilaurin by Ferstl occurs [10]. During the pressure reduction hysteresis of sound velocity and volume, changes have been observed [19]. This phenomenon reflects the coexistence of two phases during the decompression.

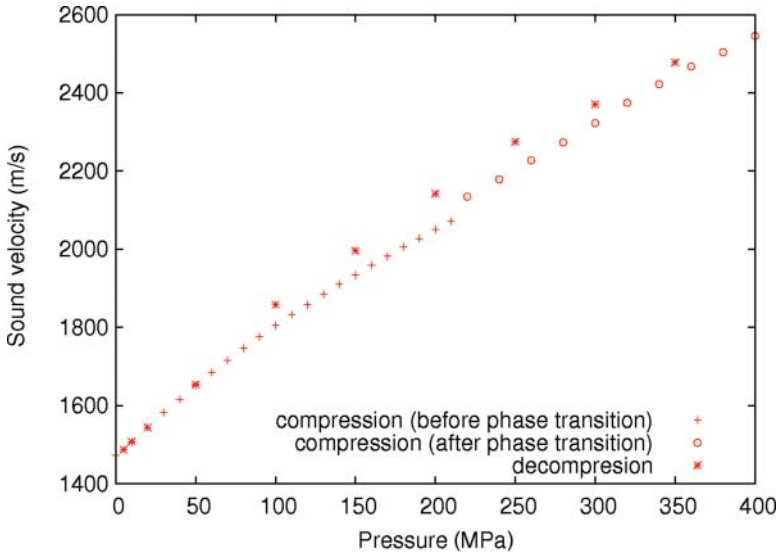


Figure 1. Sound velocity in the DAG sample in increasing and decreasing pressure.

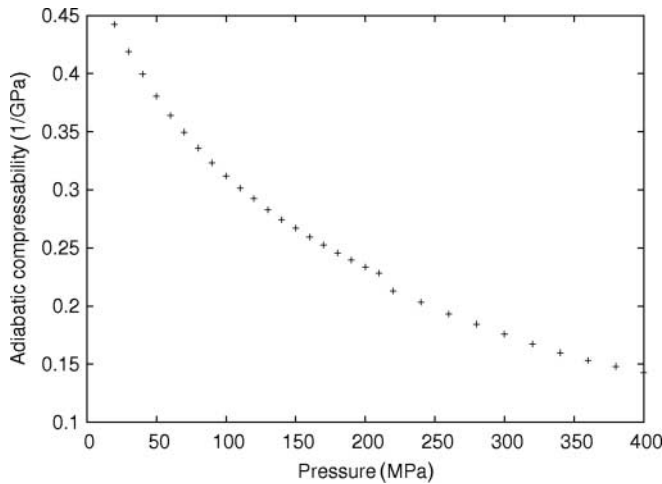


Figure 2. Adiabatic compressibility versus pressure.

The density, and adiabatic compressibility were evaluated. The dependence of adiabatic compressibility on pressure is shown in Figure 2. As could be noticed, the compressibility after the phase transition is much smaller. The dependence on pressure seems to be weaker.

The elastic properties of materials can also be found from the analysis of sound velocity [31,32]. For this purpose the dependence:

$$v = \sqrt{\frac{K}{d}},$$

where K is the bulk modulus, d the density and v the compressional sound velocity, should be used for liquids whereas for solids it should be replaced by

$$v = \sqrt{\frac{E}{d}},$$

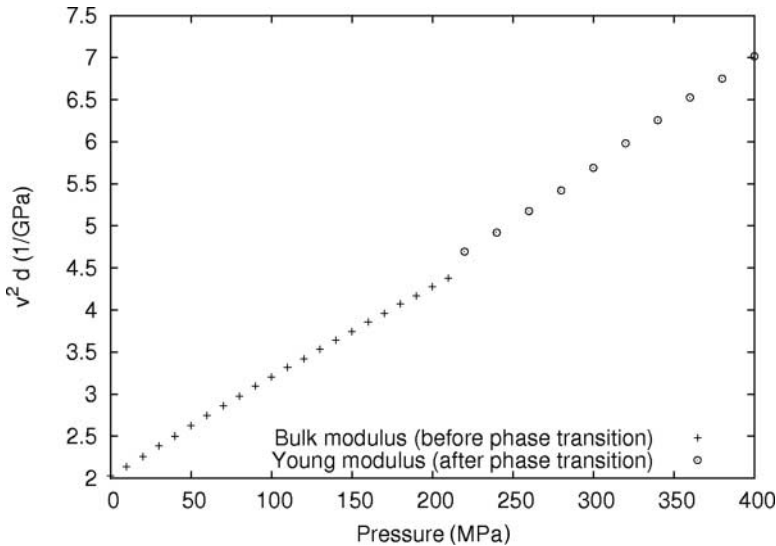


Figure 3. Bulk modulus (liquid state) marked by crosses and Young modulus (solid state) marked by open circles.

where E is the Young modulus. Thus, for both phases, the factor $v^2 d$ could be calculated. This factor is equal to K for liquids and to $E = K_s + 4G/3$ for the crystal state, where G is the shear modulus.

Bulk modulus (for liquid) and Young modulus (for crystal state) are shown in Figure 3.

5. Conclusions

- (1) Both volume on pressure and sound velocity on pressure dependence have confirmed the existence of phase transition in DAG oil.
- (2) The transition pressure is much lower than in TAGs [33]. Molecular differences between both components should be considered as the main reason of the transition's pressure divergence.
- (3) The adiabatic compressibility is clearly higher before the phase change. After the change, the compressibility has a weaker dependence on pressure. This trend is visibly reflected by the bulk and Young modulus.
- (4) During the pressure reduction hysteresis of sound velocity and volume changes has been observed. This phenomenon reflects the coexistence of two phases during the decompression.

References

- [1] Siegoczyński RM, Jedrzejewski J, Wiśniewski R. Long time relaxation effect of liquid castor-oil under high pressure conditions. *High Pressure Res.* 1989;1:225–233.
- [2] Rostocki AJ, Wiśniewski R, Wilczyńska T. High pressure phase transition in rapeseed oil. *J Mol Liq.* 2007;135:120–122.
- [3] Rostocki AJ, Kościeszka R, Tefelski DB, Kos A, Siegoczyński RM, Chruściński Ł. Pressure induced phase transition in soy oil. *High Pressure Res.* 2007;27:43–46.
- [4] Wiśniewski R, Siegoczyński RM, Długosz A, Przewłocki M, Szymański M, Trzeciecki M. Investigations of triolein under high pressure. *High Temperatures – High Pressures.* 2001;33:231–236.
- [5] Werner M, Baars A, Eder C, Delgado A. Thermal conductivity and density of plant oils under, high pressure. *J Chem Eng Data.* 2008;53(7):1444–1452.
- [6] Acosta GM, Smith RL Jr, Arai K. High-pressure PVT behavior of natural fats and oils, trilaurin, triolein, and n-tridecane from 303 K to 353 K from atmospheric pressure to 150 MPa. *J Chem Eng Data.* 1996;41:961–969.

- [7] Guignon B, Aparicio C, Sanz PD. Volumetric properties of sunflower and olive oils at temperatures between 15 and 55 C under pressures up to 350 MPa. *High Pressure Res.* 2009;29:38–45.
- [8] Rostocki AJ, Tefelski DB, Ptasznik S. Compressibility studies of some vegetable oils up to 1 GPa. *High Pressure Res.* 2009;29:721–725.
- [9] Delgado A, Kulisiewicz L, Rauh C, Benning R. Basic aspects of phase changes under high pressure. *Ann NY Acad Sci.* 2010;1189:16–23.
- [10] Ferstl P, Eder C, Russ W, Wierschem A. Pressure-induced crystallization of triacylglycerides. *High Pressure Res.* 2011;31:339–349.
- [11] Przedmojski J, Siegoczyński RM. X-ray diffraction investigation of oleic acid under high pressure. *Phase Trans.* 2002;75:581–585.
- [12] Kościeszka R, Kulisiewicz L, Delgado A. Observations of a high-pressure phase creation in oleic acid. *High Pressure Res.* 2010;30:118–123.
- [13] Kościeszka R, Siegoczyński RM, Rostocki AJ, Tefelski DB, Kos A, Ejchart W. Relative permittivity behavior and temperature changes in linoleic acid during phase transition. *J Phys Conf Ser.* 2008;121:142005.
- [14] Kos A, Siegoczyński RM, Rostocki AJ, Tefelski DB, Kościeszka R, Wieja K. Multiple phase transitions in palmitoleic acid under the pressure. *J Phys Conf Ser.* 2008;121:142008.
- [15] Rostocki AJ, Siegoczyński RM, Kielczyński P, Szalewski M, Balcerzak A, Zduniak M. Employment of a novel ultrasonic method to investigate high pressure phase transitions in oleic acid. *High Pressure Res.* 2011;31:334–338.
- [16] Kościeszka R, Tefelski DB, Ptasznik S, Rostocki AJ, Malanowski A, Siegoczyński RM. A study of the high pressure phase transition of diacylglycerol oil by means of light transmission and scattering. *High Pressure Res.* 2012;32:323–329.
- [17] Kielczyński P, Szalewski M, Balcerzak A, Malanowski A, Siegoczyński RM, Ptasznik S. Investigation of high pressure phase transitions in DAG (diacylglycerol) oil, using the Bleustein-Gulyaev ultrasonic wave method. *Food Res Int.* 2012;49:60–64.
- [18] Siegoczyński RM, Rostocki AJ, Kielczyński P, Szalewski M. A viscosity measurement during the high pressure phase transition in triolein. *J Phys Conf Ser.* 2008;121:142010.
- [19] Rostocki AJ, Siegoczyński RM, Kielczyński P, Szalewski M, Balcerzak A, Zduniak M. Employment of a novel ultrasonic method to investigate high pressure phase transitions in oleic acid. *High Pressure Res.* 2011;31:332–338.
- [20] Kielczyński P, Szalewski M, Rostocki AJ, Zduniak M, Siegoczyński RM, Balcerzak A. Investigation of high-pressure phase transitions in vegetable oil by measuring phase velocity of longitudinal ultrasonic waves. *IEEE International Ultrasonics Symposium Proceedings*; 2009; Rome, Italy; 1563–1566.
- [21] Kaneko F, Yano J, Sato K. Diversity in the fatty-acid conformation and chain packing of cis unsaturated lipids. *Curr Opin Struct Biol.* 1998;8:417–425.
- [22] Sato K. Solidification and phase transformation behaviour of food fats – a review. *Fett/Lipid.* 1999;101:467–474.
- [23] Lo SK, Tan CP, Long K, Yusoff MSA, Lai OM. Diacylglycerol oil – properties, processes and products. A review. *Food Bioprocess Technol.* 2008;1:223–233.
- [24] Nagao T, Watanabe H, Goto N, Onizawa K, Taguchi H, Matsuo N, Yasukawa T, Tsushima R, Shimasaki H, Itakura H. Dietary diacylglycerol suppresses accumulation of body fat compared to triacylglycerol in men in a double-blind controlled trial. *J Nutr.* 2000;130:792–797.
- [25] Teramoto T, Watanabe H, Ito K, Omata Y, Furukawa T, Shimoda K, Hoshino M, Nagao T, Naito S. Significant effects of diacylglycerol on body fat and lipid metabolism in patients on hemodialysis. *Clin Nutr.* 2004;23:1122–1126.
- [26] Taguchi H, Omachi T, Nagao T, Matsuo N, Tokimitsu I, Itakura H. Dietary diacylglycerol suppresses high fat diet-induced hepatic fat accumulation and microsomal triacylglycerol transfer protein activity in rats. *J Nutr Biochem.* 2002;13:678–683.
- [27] Wierzbiński M, Kościeszka R, Tefelski DB, Siegoczyński R. Determination of thermodynamic parameters of oleic acid under high pressure. *High Pressure Res.* 2010;30:135–141.
- [28] Rostocki AJ, Urbański M, Wiśniewski R, Wilczyńska T. On the improvement of the metrological properties of manganin sensors. *Metrologia.* 2005;42:250–252.
- [29] Rostocki AJ, Wiśniewski R. Linear unbalanced dc bridge. *Rev Sci Inst.* 1977;75:710–711.
- [30] Sugawara S. Time difference measurement of ultrasonic pulses using cross-correlation function between analytic signal. *Jap J Appl Phys.* 2002;41:3299–3307.
- [31] Coupland JN, McClements DJ. Physical properties of liquid edible oils. *J Am Oil Chem Soc.* 1997;74:1559–1564.
- [32] Sankarappa T, Kumar MP, Ahmad A. Ultrasound velocity and density studies in some refined and unrefined edible oils. *Phys Chem Liq.* 2005;43:507–514.
- [33] Rostocki AJ, Siegoczyński RM, Kielczyński P, Szalewski M. An application of love SH waves for the viscosity measurement of triglycerides at high pressures. *High Pressure Res.* 2010;30:88–92.